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From Tri-O-Acetyl-D-Glucal to (2R,3R,5R)-2,3-Diazido-5-

Hydroxycyclohexanone Oxime

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From Tri-O-Acetyl-D-Glucal to (2R,3R,5R)-2,3- Diazido-5-Hydroxycyclohexanone Oxime

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CONTENTS

ABSTRACT

Methyl 3-azido-2,3-dideoxy- α/β -D-arabino- and $-\alpha/\beta$ -D-ribo-hexopyranosides were transformed into 6-iodo analogues via p -tolylsulfonyl compounds. Elimination of hydrogen iodide from 6-iodo glycosides provided methyl 4-O-acetyl-3-azido-2,3,

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6-trideoxy- α - and - β -D-threo-hex-5-eno-pyranosides or 3-azido-4-O-p-tolylsulfonyl- $2,3,6$ -trideoxy- α -D-threo- and - β -D-erythro-hex-5-eno-pyranosides. Ferrier's carbocyclization of 4-O-acetyl-3-azido-2,3,6-trideoxy- α - and - β -D-threo-hex-5-eno-pyranosides gave (2S,3R,5R)-2-acetoxy-3-azido-5-hydroxycyclohexanone, which was converted into oxime. The 2-OAc group in oxime was substituted by azide ion to yield (2R,3R,5R)-2,3 diazido-5-hydroxycyclohexanone oxime. The configuration and conformation of all products are widely discussed on the basis of the ${}^{1}H$ and ${}^{13}C$ NMR.

Key Words: Azide; Tosylation; Iodination; Hex-5-eno-pyranoside; Ferrier's carbocyclization; Oxime.

INTRODUCTION

Ferrier's carbocyclization reaction is one of the most efficient methods for the preparation of chiral substituted cyclohexanones from aldohexoses.[1,2] Products of this reaction are used as precursors of different bio-active substances such as cyclitols and aminocyclitols $[3 - 10]$ as well as natural compounds and their analogues containing a cyclitol unit.^[11-20] Previously our interest was focused on the addition of hydrazoic acid to α , β -unsaturated aldehydes derived from acetylated glycals, a useful source of 3-azido-2,3-dideoxy sugars.^[21-23] In this paper, we report the preparation of 2-acetoxy-3-azido-5-hydroxycylohexanone oxime from tri-O-acetyl-D-glucal via addition of hydrazoic acid followed by Ferrier's carbocyclization. Combination of this addition with Ferrier's carbocyclization could be a simple procedure of obtaining amino- and diaminocyclitols. Bearing in mind our previous experiences concerning 2-hydroxyimino glycosides, $[24-26]$ we explored the ability of substitution of 2-OAc group by azide ion in this cyclohexanone oxime.

RESULTS AND DISCUSSION

Fourstereoisomersofmethyl3-azido-2,3-dideoxyhexopyranosides(1–4,Schs.1and 2) were synthesized starting from tri-O-acetyl-D-glucal as previously reported.^[22] In order to transform them into methyl 3-amino-6-iodo-2,3,6-trideoxyhexopyranosides, first Garegg's method $[I_2, \text{imidazole}, P(\text{Ph})_3]$ was employed.^[27] We assumed that triphenylphosphine, used in iodination procedure, would also reduce $3-N_3$ group. This idea was abandoned because the only product formed during reaction course was identified as methyl 3-amino-2,3-dideoxyhexopyranoside (Sch. 1). No iodination reaction occurred, no matter whether the reaction was carried out at room or at higher temperatures (70° C) and 120° C). The same 3-amino-2,3-dideoxyhexopyranoside was obtained by direct reduction of $3-N_3$ group when triphenylphosphine or catalytic hydrogenation was used. Because Garegg's method was applied successfully with other compounds, $[3,16,17]$ we concluded that $3-N_3$ group, present in our substrates, is responsible for the lack of the iodination reaction.

Similarly, iodination products were not detected while Anisuzzaman's method [CI4, $P(Ph)_{3}$, pyridine] was applied to 3-azido glycosides (Sch. 1).^[28]

Iodination of methyl 3-acetamido-2,3-dideoxyhexopyranosides, derived from 3-amino analogues (Sch. 1) with Garegg's or Anisuzzaman's strategy, was also unsuccessful.

These negative results prompted us to introduce iodide ion *via* tosylation reaction of the terminal hydroxyl group. Tosylation of $1-4$ with p-tolylsulfonyl chloride in pyridine (Sch. 2) gave respective mixtures of methyl 3-azido-2,3-dideoxy-6- O -p-tolylsulfonyl- (5, 7, 9) and -4,6-di-O-p-tolylsulfonylhexopyranosides (6, 8, 10, 11). Contrary to our expectation, tosylation reaction was not regioselective, thus both the primary and secondary hydroxyl groups were substituted. In the case of 4 only ditolylsulfonyl derivative 11 was gained.

The presence of the tosyl groups in $5-11$ is undoubtedly proved by IR, ¹H, and ¹³C NMR data. Worthy of notice is a significant difference in the chemical shifts of H-4

protons in 5–11 when monotosyl (4-OH) and ditosyl (4-OTs) analogues are compared $(\Delta \delta 0.8-0.9)$, showing the deshielding influence of the *p*-tolylsulfonyl group (Table 1).

Acetylation of monotosyl glycosides 5, 7, and 9 led to 12, 13, and 14, which were refluxed with sodium iodide in acetone to yield methyl 4-O-acetyl-3-azido-6-iodo-2,3,6 trideoxyhexopyranosides (15, 16, and 17, respectively). The same iodination reaction of ditosyl glycosides (6, 10, and 11) was regioselective and gave only methyl 3-azido-6 iodo-4-O-p-tolylsulfonyl-2,3,6-trideoxyhexopyranosides (18, 19, and 20, respectively). The signals of the protons of p -tolylsulfonyl group were absent in the ${}^{1}H$ NMR spectra of 15–17 and corresponded to one tosyl group in 18–20. Additionally, the H-6 protons signals of 6-iodo compounds (15–20) were shifted to a higher field by ~ 0.9 ppm as compared to H-6 protons of 6-OTs glycosides (5–14), indicating replacement of the p-tolylsulfonyl group. Noteworthy is the relatively small chemical shift of C-6 carbon in the ¹³C NMR spectra of **15–20** (δ 4–6), which we found characteristic for 6-iodo sugars (Table 3).

The coupling constants of glycosides 5–20 confirm their structures (Table 2). Thus, the coupling constants $J_{2a,3} \sim 12 \text{ Hz}$ and $J_{3,4} \sim J_{4,5} = 9-10 \text{ Hz}$ are indicative for D-arabino (5–8, 12, 13, 15, 16, 18) while $J_{2a,3} \sim J_{3,4} = 3-4$ Hz and $J_{4,5} = 9-10$ Hz for D -ribo configurations (9-11, 14, 17, 19, 20). All anomeric protons usually appear as doublets with coupling constant $J_{1,2a} = 3-4 Hz$ (α anomers) or $8-9 Hz$ (β anomers). The $J_{1,2e}$ coupling constant is not recorded or is small $(1-2 Hz)$ for 2-deoxyglycosides $5-20$, which is due to the anti-periplanar orientation of the ring oxygen atom to equatorially oriented $H-2_e$ proton.^[29] Additional proof for configuration of the anomeric carbon comes from the chemical shifts of H-1 protons. The H-1 signals of α anomers (5, 6, 9, 10, 12, **14, 15, 17, 18, 20**) appear at higher δ values than those of the analogous protons of β anomers (7, 8, 11, 13, 16, 19), owing to the respective equatorial and axial orientation of the H-1 proton (Table 1).

All the aforementioned findings are in accordance with the ${}^{4}C_{1}$ (D) conformation of compounds 5–20.

6-Iodo glycosides $(15-20)$ treated with silver fluoride in pyridine gave methyl 4-O-acetyl-3-azido- (21, 22) and -3-azido-4-O-p-tolylsulfonyl-2,3,6-trideoxy-hex-5-enopyranosides (23, 24), respectively. Lack of both, the H-5 proton signal and geminal coupling constant $J_{6,6'} = 10-14$ Hz, in the ¹H NMR spectra of 21-24 is an evidence for 5-eno structure of 21–24. Next, the chemical shifts of the H-6 protons ($\delta \sim 4.7$) show deshielding influence of the double bond, which is also observed in the 13 C NMR spectra by the chemical shifts of the C-5 and C-6 carbons (\sim 150 and \sim 100 ppm, respectively).

Introduction of an exocyclic double bond to the pyranose ring has a significant influence on the conformation of 5-enoglycosides (21–24, Sch. 3). Only these with α -D-threo configurations (21 and 23) keep the 4C_1 form, which is demonstrated by coupling constants $J_{2a,3} \sim 12$ Hz and $J_{3,4} \sim 10$ Hz. The 4C_1 conformation is optimal for 21 and 23 because both $3-N_3$ and $4-OAc$ groups are equatorially oriented and an anomeric effect is omitted. The opposite configuration of the anomeric carbon is the sole difference when 21 (α -D-threo) and 22 (β -D-threo) are compared. This difference causes a deviation from ${}^{4}C_1$ form in the case of 22, probably due to an unfavorable anomeric effect. The coupling constants $J_{2a,3} = J_{3,4} = 8.1$ Hz and $J_{1,2a} = 5.8$ Hz allow us to presume the conformational equilibrium between ${}^{4}C_{1}$ and ${}^{1}C_{4}$ forms in 22. Both an anomeric effect and axially oriented 3-N₃ group are responsible for the change of the ${}^{4}C_{1}$ into ${}^{1}C_{4}$ form in 24. The ¹C₄ conformation of 24 is established on the basis of $J_{1,2a} = 3.5$ Hz and

Table 1. The chemical shifts in ¹H NMR spectra (ppm) for compounds 5-24 (400 MHz, CDCl₃). Table 1. The chemical shifts in 1H NMR spectra (ppm) for compounds 5–24 (400 MHz, CDCl3).

| | Configuration | $J_{1,2a}$ | $J_{1,2e}$ | $J_{2a,2e}$ | $J_{2a,3}$ | $J_{2e,3}$ | $J_{3,4}$ | $J_{4,5}$ | $J_{5,6}$ | $J_{5,6'}$ | $J_{6,6'}$ |
|----|---------------------|------------|------------|-------------|------------|------------|-----------|-----------|-----------|------------|------------|
| 5 | α -D-arabino | 3.4 | | 13.5 | 12.5 | 5.3 | 9.6 | 9.6 | 4.3 | 1.9 | 11.1 |
| 6 | α -D-arabino | 3.3 | | 13.1 | 12.3 | 5.7 | 10.2 | 9.4 | 6.5 | 2.0 | 11.1 |
| 7 | β -D-arabino | 9.0 | | 12.9 | 12.4 | 3.4 | 9.6 | 9.6 | 3.9 | | 11.3 |
| 8 | β -D-arabino | 9.2 | 1.8 | 12.9 | 12.0 | 3.4 | 9.2 | 9.2 | 6.5 | 2.8 | 11.1 |
| 9 | α -D-ribo | 3.8 | | 14.7 | 4.3 | 2.4 | 3.3 | 9.9 | 4.3 | | 11.4 |
| 10 | α -D-ribo | 4.0 | | 14.9 | 4.0 | 2.0 | 2.9 | 10.0 | 4.9 | 2.0 | 10.9 |
| 11 | β -D-ribo | 8.4 | 1.6 | 13.7 | 3.2 | 1.5 | 3.2 | 9.0 | 4.7 | 1.1 | 11.1 |
| 12 | α -D-arabino | 3.7 | | 13.2 | 12.2 | 4.7 | 9.5 | 10.1 | 5.8 | 2.6 | 10.6 |
| 13 | β -D-arabino | 9.3 | 2.1 | 13.2 | 12.3 | 4.7 | 9.8 | 9.8 | 5.9 | 3.4 | 10.6 |
| 14 | α -D-ribo | 3.8 | | 14.9 | 3.8 | 2.9 | 3.8 | 9.6 | 3.9 | 1.9 | 12.0 |
| 15 | α -D-arabino | 3.4 | 1.5 | 13.5 | 12.4 | 5.1 | 10.1 | 9.6 | 9.0 | 2.8 | 11.3 |
| 16 | β -D-arabino | 9.2 | 1.0 | 12.7 | 12.7 | 4.6 | 9.7 | 9.2 | 9.2 | 2.0 | 10.7 |
| 17 | α -D-ribo | 4.0 | 1.5 | 15.0 | 4.0 | 3.3 | 3.7 | 9.5 | 7.3 | 2.6 | 10.6 |
| 18 | α -D-arabino | 2.9 | | 13.2 | 12.4 | 5.1 | 9.5 | 9.5 | 8.8 | 2.6 | 11.0 |
| 19 | β -D-ribo | 9.0 | 2.0 | 14.0 | 3.5 | 4.0 | 3.5 | 9.0 | 8.0 | 3.5 | 11.0 |
| 20 | α -D-ribo | 4.5 | | 14.5 | 4.5 | 2.7 | 3.6 | 9.0 | 8.1 | 2.7 | 10.9 |
| 21 | α -D-threo | 3.3 | 1.5 | 13.6 | 12.1 | 4.8 | 9.9 | | | | 1.5 |
| 22 | β -D-threo | 5.8 | 3.5 | 13.9 | 8.1 | 5.2 | 8.1 | | | | |
| 23 | α -D-threo | 3.3 | 1.8 | 13.6 | 11.7 | 4.8 | 10.2 | | | | |
| 24 | β -D-erythro | 3.5 | 1.0 | 13.0 | 13.0 | 4.5 | 3.0 | | | | |

Table 2. The ${}^{1}H$ - ${}^{1}H$ coupling constants (Hz) for compounds $5-24$ (400 MHz).

 J_{2a} = 13.0 Hz coupling constants, which are characteristic for equatorial (H-1) and axial $(H-2_a$ and H-3) orientation of the respective protons. These findings are in accordance with the findings of the Sztaricskai group.^[10]

As demonstrated, the conformation of the 5-eno glycosides $(21-24)$ is flexible, and even a small change in their configuration has a significant influence on their conformation. This can be explained by an $sp²$ hybridization of the C-5 carbon, which causes no significant differences between ${}^{4}C_{1}$ and ${}^{1}C_{4}$ forms. If the $-CH_{2}OR$ group is bonded to an sp³ hybridized C-5 carbon atom, its equatorial orientation is strongly preferred and forces the 4C_1 (D) form.

The mercury salt mediated carbocyclic ring transformation reaction of 21 and 22 led to $(2S,3R,5R)$ -2-acetoxy-3-azido-5-hydroxycyclohexanone (25) . As expected,^[10,30] carbocyclization was highly stereoselective and provided only cyclohexanone with 5-OH and $3-N_3$ groups *trans*. The structure of 25 was established as follows. Instead of one, there are two methylene groups in the ${}^{1}H$ NMR spectrum of 25 with chemical shifts at 1.91 $(H-4_a)$, 2.42 $(H-4_e)$, 2.64 $(H-6_a)$, and 2.68 $(H-6_e)$ (Table 4). Presence of the 5-OH group is demonstrated by the signal at 3.75 ppm and by the IR spectra, where OH band (3467 cm^{-1}) is recorded. In the ¹³C NMR spectra the C-1 carbon atom is the most dishielding (δ 199.08), indicating ketone carbonyl group (Table 6). The crucial question in the structure of 25 refers to the configuration of the newly generated asymmetric center (C-5). The coupling constants $J_{2,3} = 10.5$ and $J_{3,4a} = 12.5$ Hz point to the axial orientation of H-2, H-3, and H-4_a protons and the ²C₅ conformation of 25. The ²C₅ form, together with coupling constants $J_{4a,5} = 3.5$ Hz, $J_{4e,5} = 2.0$ Hz, and $J_{5.6a} = J_{5.6e} = 3.0$ Hz, indicate 5R configuration of C-5 carbon atom (Table 5).

Table 3. The chemical shifts in ¹³C NMR spectra (ppm) for compounds 5-24 (400 MHz). **Table 3.** The chemical shifts in 13 C NMR spectra (ppm) for compounds $5-24$ (400 MHz).

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| | $H-2$ | $H-3$ | $H-4a$ | $H-4$ | $H-5$ | $H-6a$ | $H-6e$ | $2-OAc$ | $5-OH$ | NOH |
|-------------------|-------|--------------------------|--------------------------|-------|-----------------------|--------|--------|---------|--------|------------|
| 25 | 5.18 | 4.22 | 1.91 | 2.42 | 4.50 | 2.64 | 2.68 | 2.24 | 3.75 | |
| CDCl ₃ | (d) | $\left(\text{qd}\right)$ | $\left(\text{qd}\right)$ | (dq) | $\left($ qw $\right)$ | (dd) | (dd) | (s) | (bs) | |
| 26 | 5.16 | 4.06 | 1.84 | 1.97 | 3.91 | 2.41 | 2.76 | 2.09 | 5.01 | 11.00 |
| DMSO | (d) | (td) | (m) | (m) | (oct) | (dd) | (dd) | (s) | (d) | (s) |
| 27 | 4.43 | 3.98 | 2.09 | 2.30 | 4.01 | 2.52 | 3.27 | | 5.27 | \pm |
| CDCl ₃ | (d) | (m) | (m) | (m) | (m) | (dd) | (dd) | | (b) | |
| 28 | 4.01 | 4.02 | 1.85 | 2.25 | 4.28 | 2.55 | 3.07 | | 5.00 | \approx |
| CDCl ₃ | (d) | (m) | (dq) | (dq) | (m) | (dd) | (dd) | | (b) | |

Table 4. The chemical shifts in ¹H NMR spectra (ppm) for compounds 25, 26 (400 MHz) and 27, 28 (100 MHz).

Not determinated.

Reaction of ketone 25 with hydroxylamine yielded the oxime 26. The transformation of carbonyl into hydroxyimino group is proved by the signal of N-OH proton in the ¹H NMR spectrum of 26 in DMSO solution (δ 11.00) and by the change in chemical shifts of the C-1 carbon atom in the 13 C NMR spectrum (Table 6). The signal of oxime carbon atom in 26 is shifted to the higher field $(\delta 150.15)$ in comparison to the carbonyl carbon atom in 25 (δ 199.08). The configuration of 26 remains the same as of 25. However, a new stereogenic center resulting from an oxime group was created. Our investigations in NOESY experiment showed the interactions between N-OH and equatorially oriented H-6e protons and, consequently, the lack of the interactions between N-OH and H-2 protons. This means that oxime 26 has E configuration, which should be favorable because the equatorially oriented 2-OAc group in 26 is a steric hindrance for the Z orientation of hydroxyimine group.

It was reported that some groups neighboring to hydroxyimino function are easily eliminated to cause their replacement by nucleophiles including azide ion.^[24-26] Thus, reaction of oxime 26 with sodium azide in ethanol provided (Z) and (E) $(2R,3R,5R)$ -2,3-diazide-5-hydroxycyclohexanone oximes (27 and 28, respectively). Lack of the carbonyl group vibrations in the IR spectra of 27 and 28 (1760 cm⁻¹ for 26) indicates the substitution of 2-OAc group by azide ion. Additionally, the chemical shift of H-2 proton of starting oxime 26 is shifted to the lower fields when compared with H-2 proton signals of 27 and 28 ($\Delta \delta$ 0.73 and 1.15, respectively), which is due to the change of the 2-OAc into 2-N₃ group.^[23] The $J_{5,6a} = 2.5$ and 3.5 Hz coupling constants

Table 5. The ${}^{1}H$ - ${}^{1}H$ coupling constants (Hz) for compounds 25, 26 (400 MHz) and 27, 28 (100 MHz).

| | | $J_{2,3}$ $J_{3,4a}$ $J_{3,4e}$ $J_{4a,4e}$ $J_{4a,5}$ $J_{4e,5}$ $J_{5,6a}$ $J_{5,6e}$ $J_{5,OH}$ $J_{6a,6e}$ | | | | |
|----|----------------|--|--|---------|-----|----------|
| | | 25 10.5 12.5 5.0 14.4 3.5 2.0 3.0 3.0 * 14.4 | | | | |
| | 26 8.1 9.3 3.9 | 13.2 2.9 | | 5.8 3.7 | 5.6 | 3.4 14.4 |
| | | 27 3.0 10.5 3.5 14.0 3.0 * 2.5 3.0 * 15.0 | | | | |
| 28 | | 3.0 11.5 5.0 13.0 3.0 2.0 3.5 5.5 * 15.0 | | | | |

Not determinated.

| | $C-1$ | $C-2$ | $C-3$ | | | | C-4 C-5 C-6 CH ₃ (Ac) C=O(Ac) | |
|----|--------|-------|-------|-------|-------------|-------|--|--------|
| 25 | 199.08 | 80.70 | 59.39 | 36.80 | 65.96 47.49 | | 20.92 | 169.87 |
| 26 | 150.15 | 72.84 | 58.76 | 35.14 | 63.16 | 30.55 | 20.65 | 169.13 |

Table 6. The chemical shifts in ¹³C NMR spectra (ppm) for compounds 25 and 26 (400 MHz).

preclude the ⁵ C_2 form for 27 and 28; thus, relatively small $J_{2,3} = 3$ Hz coupling constant results from equatorial orientation of H-2 proton in 27 and 28, which adopt ${}^{2}C_{5}$ form. This means that the configuration of $C-2$ carbon atom changed during reaction course and is R now. Discussed reaction occurs via elimination–addition mechanism, which probably involves a reactive intermediate 29 (Sch. 3)^[24] and thus the change of C-2 as well as oxime configurations seems to be possible. The oxime and neighboring equatorial groups are nearly coplanar placed; therefore, the azide group prefers axial orientation in newly generated oximes 27 and 28. This arrangement avoids unfavorable steric and electrostatic interactions. Bearing in mind previous conclusions concerning the effect of the hydroxyimino group on the position of ${}^{1}H$ NMR signals of adjacent protons^[31,32] as well as the chemical shifts of H-2 for 27 (δ 4.43) and 28 (δ 4.01), it is established that oxime group has the Z orientation in 27 and E orientation in 28.

EXPERIMENTAL

General methods. Melting points are uncorrected. Optical rotations were recorded at room temperature (20° C) using a Hilger-Watt polarimeter for solutions in CHCl₃. TLC was performed on the Merc Kieselgel 60 F-254 plates with: A, petroleum ether–AcOEt (1 : 2); B, petroleum ether–AcOEt (2 : 1); C, petroleum ether–AcOEt (4 : 1); D, CCl₄– acetone $(3:1)$; E, n-heptane–AcOEt $(1:2)$; F, n-heptane–AcOEt $(1:1)$; G, CHCl₃– AcOEt (1 : 2); H, CHCl₃ $-$ AcOEt (2 : 1). Column chromatography was performed on MN Kieselgel 60 (<0.08 mm). The ¹H and ¹³C NMR spectra (CDCl₃ or DMSO, internal Me4Si) were recorded with a Varian Mercury 400 (400.49/100.70 MHz) or Varian XL-100 (100 MHz) instruments. IR spectra were recorded as Nujol mulls with a Bruker IFS 66 spectrophotometer. Elemental analyses were conducted with a Carlo Erba EA1108 elemental analyzer.

Methyl 3-azido-2,3-dideoxy- α -D-arabino- (1), - β -D-arabino- (2), - α -D-ribo- (3), and $-\beta$ -D-ribo-hexopyranosides (4). Prepared according to procedure previously reported.[22]

General procedure for tosylation. To the solutions of $1-4$ (0.2 g, 1 mmol) in CH_2Cl_2 (10 mL), dry pyridine (0.5 mL) and p-toluenesulfonyl chloride (0.95 g, 5 mM) were added. The mixtures were stirring at rt for 24–72 h, depending on the substrate. The end of the reactions was detected by TLC (solvent A). Then the mixtures were diluted with Et_2O (10 mL). Precipitated salts were filtered off. The filtrates were concentrated and diluted with CHCl₃. The organic solutions were washed with satd NaHCO₃ solution, and with water and dried over $Na₂SO₄$. Concentration under reduced pressure led to the crude products, which were chromatographed (solvent B).

Methyl 3-azido-2,3-dideoxy-6-O-p-tolylsulfonyl- (5) and -4,6-di-O-p-tolylsulfonyl- α -D-arabino-hexopyranosides (6). Tosylation of 1 yielded a mixture of two products, which were separated by column chromatography to give 5 and 6, reported previously.[22]

Methyl 3-azido-2,3-dideoxy-6-O-p-tolylsulfonyl- (7) and -4,6-di-O-p-tolylsulfonyl- β -D-arabino-hexopyranosides (8). Tosylation of 2 yielded a mixture of two products, which were separated by column chromatography to give first 7 (77%, syrup); $\lbrack \alpha \rbrack_{0}^{20}-3^{\circ}$ (c 0.8, CHCl₃); R_f 0.45 (solvent B); IR: ν 3511 (OH), 2103 (N₃), 1597 (C=C_{ar}), 1360, 1190, 1176 (O=S=O) cm⁻¹.

Anal. Calcd for $C_{14}H_{19}N_3O_6S$: C 47.05, H 5.35, N 11.76, S 8.97; Found: C 47.83, H 5.48, N 11.11, S 8.99.

Eluted second was 8 (2%); mp 119-121°C; $[\alpha]_D^{20} + 44^\circ$ (c 0.2, CHCl₃); R_f 0.52 (solvent B); IR: ν 2106 (N₃), 1597 (C=C_{ar}), 1362, 1190, 1176 (O=S=O) cm^{-1} .

Anal. Calcd for $C_{21}H_{25}N_3O_8S_2$: C 49.30, H 4.93, N 8.21, S 12.54; Found: C 49.75, H 4.97, N 8.17, S 11.95.

Methyl 3-azido-2,3-dideoxy-6-O-p-tolylsulfonyl- (9) and -4,6-di-O-p-tolylsulfonyl- α -D-ribo-hexopyranosides (10). Tosylation of 3 yielded a mixture of two products, which were separated by column chromatography to give first 9 (53%, syrup); $[\alpha]_D^{20} + 177^\circ$ (c 0.3, CHCl₃); R_f 0.31 (solvent B); IR: ν 3496 (OH), 2105 (N₃), 1597 (C=C_{ar}), 1357, 1175 $(O=$ S=O) cm⁻¹.

Anal. Calcd for $C_{14}H_{19}N_3O_6S$: C 47.05, H 5.35, N 11.76, S 8.97; Found: C 48.50, H 5.31, N 10.68, S 9.31.

Eluted second was 10 (18%, syrup); $[\alpha]_D^{20} + 125^\circ$ (c 1.2, CHCl₃); R_f 0.47 (solvent B); IR: v 2107 (N₃), 1597 (C=C_{ar}), 1365, 1190, 1176 (O=S=O) cm⁻¹.

Anal. Calcd for $C_{21}H_{25}N_3O_8S_2$: C 49.30, H 4.93, N 8.21, S 12.54; Found: C 50.77, H 5.34, N 7.90, S 11.50.

Methyl 3-azido-2,3-dideoxy-4,6-di-O-p-tolylsulfonyl- β -D-ribo-hexopyranoside (11). Tosylation of 4 yielded a crude product, which was chromatographed to give 11 (62%); mp 99–101°C; $[\alpha]_D^{20}$ +26° (c 0.6, CHCl₃); R_f 0.62 (solvent B); IR: v 2104, (N₃), 1597 (C=C_{ar}), 1363, 1190, 1176 (O=S=O) cm⁻¹.

Anal. Calcd for $C_{21}H_{25}N_3O_8S_2$: C 49.30, H 4.93, N 8.21, S 12.54; Found: C 49.46, H 4.92, N 7.88, S 12.32.

General procedure for acetylation. Compounds $5, 7$, and 9 (0.357 g, 1 mmol) were acetylated with $Ac_2O(2 \text{ mL})$ and pyridine (2 mL), respectively. During 0.5 h the reactions were over (TLC, solvent B). After dilution with CH_2Cl_2 (10 mL) the organic solutions were washed with satd NaHCO₃ solution and with water and dried over Na₂SO₄. Concentration under reduced pressure led to the crude products, which were chromatographed (solvent B).

Methyl 4-O-acetyl-3-azido-2,3-dideoxy-6-O-p-tolylsulfonyl-a-D-arabino-hexo**pyranoside (12).** Acetylation of 5 led to 12, reported previously.^[22]

Methyl 4-O-acetyl-3-azido-2,3-dideoxy-6-O-p-tolylsulfonyl-β-D-arabino-hexo**pyranoside (13).** Acetylation of 7 gave 13 (92%); mp 95–97°C; $[\alpha]_D^{20} + 15^\circ$ (c 0.6, CHCl₃); R_f 0.59 (solvent B); IR: ν 2102 (N₃), 1749 (C=O), 1597 (C=C_{ar}), 1364, 1189, 1176 (O=S=O) cm⁻¹.

Anal. Calcd for $C_{16}H_{21}N_3O_7S$: C 48.11, H 5.30, N 10.52, S 8.03; Found: C 48.26, H 5.41, N 9.83, S 7.76.

Methyl 4-O-acetyl-3-azido-2,3-dideoxy-6-O-p-tolylsulfonyl-a-D-ribo-hexopyra**noside (14).** Acetylation of 9 yielded 14 (85%, syrup); $[\alpha]_D^{20} + 131^{\circ}$ (c 0.2, CHCl₃); R_f 0.56 (solvent B); IR: ν 2104 (N₃), 1742 (C=O), 1597 (C=C_{ar}), 1366, 1189, 1177 $(O=$ S=O) cm⁻¹.

Anal. Calcd for $C_{16}H_{21}N_3O_7S$: C 48.11, H 5.30, N 10.52, S 8.03; Found: C 48.69, H 5.25, N 10.01, S 7.58.

General procedure for substitution of 6-OTs group by iodide ion. The solutions of 6, 10, 11 (0.511 g, 1 mmol), and $12-14$ (0.399 g, 1 mmol) in acetone (15 mL) containing NaI (0.75 g, 5 mmol) were refluxed, respectively. After 24 h the mixtures were cooled and diluted with $Et₂O$ (10 mL). Precipitated salts were filtered off. The filtrates were concentrated and acetone (15 mL) containing NaI (0.75 g, 5 mmol) was added again. The mixtures were refluxed the next 24 h. The end of reactions was verified by TLC (solvent C). Then the mixtures were cooled and diluted with $Et₂O$ (10 mL). Precipitated salts were filtered off. The filtrates were concentrated and diluted with $CHCl₃$. The organic solutions were washed with aq $\text{Na}_2\text{S}_2\text{O}_3$ and water, dried over Na_2SO_4 , and concentrated to give the crude products, which were chromatographed (solvent C).

Methyl 4-O-acetyl-3-azido-6-iodo-2,3,6-trideoxy-a-D-arabino-hexopyranoside (15). Iodination of 12 gave 15, reported previously.^[22]

Methyl 4-O-acetyl-3-azido-6-iodo-2,3,6-trideoxy-β-D-arabino-hexopyranoside (16). Iodination of 13 yielded 16 (42%); mp 79-80°C; $[\alpha]_D^{20} + 6$ ° (c 0.4, CHCl₃); R_f 0.65 (solvent C); IR: ν 2101 (N₃), 1746 (C=O) cm⁻¹.

Anal. Calcd for C₉H₁₄N₃O₄I: C 30.44, H 3.97, N 11.83; Found: C 30.63, H 3.93, N 11.72.

Methyl 4-O-acetyl-3-azido-6-iodo-2,3,6-trideoxy- α -D-ribo-hexopyranoside (17). Iodination of 14 led to 17 (40%, syrup); $[\alpha]_D^{20} + 103^{\circ}$ (c 0.2, CHCl₃); R_f 0.58 (solvent C); IR: ν 2103 (N₃), 1739 (C=O) cm⁻¹.

Methyl 3-azido-6-iodo-4-O-p-tolylsulfonyl-2,3,6-trideoxy-a-D-arabino-hexopyranoside (18). Iodination of 6 gave 18, reported previously.^[22]

Methyl 3-azido-6-iodo-4-O-p-tolylsulfonyl-2,3,6-trideoxy- β -D-ribo-hexopyranoside (19). Reaction of 11 with NaI yielded 19 (36%); mp 78–80°C; $[\alpha]_D^{20} + 7$ ° (c 0.4, CHCl₃); R_f 0.58 (solvent C); IR: ν 2104 (N₃), 1597 (C=C_{ar}), 1371, 1190, 1177 (O=S=O) cm⁻¹.

Anal. Calcd for $C_{14}H_{18}N_3O_5SI$: C 35.99, H 3.88, N 8.99, S 6.86; Found: C 36.78, H 3.90, N 8.72, S 6.59.

Methyl 3-azido-6-iodo-4-O-p-tolylsulfonyl-2,3,6-trideoxy- α -D-ribo-hexopyranoside (20). Reaction of 10 with NaI led to 20 (38%); mp 88–90°C; $[\alpha]_D^{20} + 111^{\circ}$ (c 1.3, CHCl₃); R_f 0.39 (solvent C); IR: ν 2106 (N₃), 1597 (C=C_{ar}), 1369, 1190, 1177 $(O=$ S=O) cm⁻¹.

Anal. Calcd for C₁₄H₁₈N₃O₅SI: C 35.99, H 3.88, N 8.99, S 6.86; Found: C 37.07, H 3.92, N 8.76, S 6.86.

General procedure for elimination of hydrogen iodide. To the solutions of 15, 16 (0.284 g, 0.4 mmol) and 18, 19 (0.187 g, 0.4 mmol) in pyridine (2 mL), AgF (0.076 g, 0.6 mmol) was added, respectively. The mixtures were protected against the light and stirred. After 48 h TLC (solvent C) indicated the end of the reactions. Then the mixtures were diluted with $Et_2O(10 \text{ mL})$. Precipitated salts were filtered off. The filtrates were concentrated and diluted with CHCl₃. The organic solutions were washed with aq $\text{Na}_2\text{S}_2\text{O}_3$ and water, dried over $Na₂SO₄$, and concentrated. The crude products were chromatographed (solvent C).

Methyl 4-O-acetyl-3-azido-2,3,6-trideoxy- α -D-threo-hex-5-eno-pyranoside (21). Reaction of 15 with AgF in pyridine gave 21 (69%, syrup); $[\alpha]_D^{20} + 112^\circ$ (c 0.2, CHCl₃); R_f 0.72 (solvent C); IR: ν 2105 (N₃), 1746 (C=O), 1662 (C=C) cm⁻¹.

Anal. Calcd for C₉H₁₃N₃O₄: C 47.57, H 5.77, N 18.49; Found: C 47.96, H 5.97, N 18.06.

Methyl 4-*O*-acetyl-3-azido-2,3,6-trideoxy-β-D-threo-hex-5-eno-pyranoside (22). Reaction of 16 with AgF in pyridine yielded 22 (72%, syrup); $[\alpha]_D^{20} - 105^\circ$ (c 0.3, CHCl₃); R_f 0.60 (solvent C); IR: ν 2104 (N₃), 1749 (C=O), 1663 (C=C) cm⁻¹.

Anal. Calcd for C₉H₁₃N₃O₄: C 47.57, H 5.77, N 18.49; Found: C 47.77, H 5.93, N 17.50.

Methyl 3-azido-4-O-p-tolylsulfonyl-2,3,6-trideoxy- α -D-threo-hex-5-eno-pyranoside (23). Reaction of 18 with AgF in pyridine led to 23 (63%, syrup); $[\alpha]_D^{20} + 174^\circ$ (c 0.2, CHCl₃); R_f 0.50 (solvent C); IR: ν 2107 (N₃), 1664 (C=C), 1599 (C=C_{ar}), 1371, 1190, 1177 (O=S=O) cm⁻¹.

Methyl 3-azido-4-*O-p*-tolylsulfonyl-2,3,6-trideoxy-β-D-erythro-hex-5-eno-pyra**noside (24).** Reaction of 19 with AgF in pyridine gave 24 (68%, syrup); $[\alpha]_D^{20} + 20^\circ$ (c 0.2, CHCl₃); R_f 0.48 (solvent C); IR: ν 2105 (N₃), 1668 (C=C), 1597 (C=C_{ar}), 1365, 1190, 1177 (O=S=O) cm⁻¹.

 $(2S, 3R, 5R)$ -2-Acetoxy-3-azide-5-hydroxycyclohexanone (25) . To a solution of 21 $(0.12 \text{ g}, 0.53 \text{ mmol})$ in 1,4-dioxane (3 mL) , 5 mM H_2SO_4 (1.3 mL) and catalytic amount of $HgSO₄$ were added. The mixture was boiled under reflux. After 15 min TLC (solvent C) showed the end of the reaction. The cooled mixture was extracted with ethyl acetate $(4 \times 5 \text{ mL})$. The combined extracts were washed with aq NaHCO₃ and water, dried over $Na₂SO₄$, and concentrated. The residue was chromatographed (solvent D) to give **25** (88%); mp 80–81°C; $[\alpha]_D^{20}$ – 14° (c 1.0, CHCl₃); R_f 0.38 (solvent D); IR: v 3467 (OH), 2109 (N₃), 1744 (ester C=O), 1736 (ketone C=O) cm⁻¹.

Anal. Calcd for C₈H₁₁N₃O₄: C 45.07, H 5.20, N 19.71; Found: C 45.12, H 5.42, N 19.27.

Analogous procedure applied to 22 led to 25 in 85% yield.

(E)-(2S, 3R, 5R)-2-acetoxy-3-azide-5-hydroxycyclohexanone oxime (26). To a solution of 25 (0.09 g, 0.42 mmol) in MeOH (1.3 mL), hydroxylamine hydrochloride (0.0293 g, 0.42 mmol) and pyridine (0.16 mL) were added. The mixture was stirred at rt. After 1 h TLC (solvent E) indicated the lack of the substrate. Then the mixture was diluted with CHCl₃, extracted with H₂O, dried (Na_2SO_4) , and concentrated. Column chromatography (solvent F) of the residue yielded 26 (92%, syrup); $[\alpha]_D^{20} - 64^\circ$ (c 1.1, CHCl₃); R_f 0.52 (solvent E); IR: ν 3600 (OH), 2100 (N₃), 1760 (ester C=O), 1640 $(C=N \text{ oxime})$, 1240 (ester C-O-C) cm⁻¹.

Anal. Calcd for $C_8H_{12}N_4O_4$: C 42.11, H 5.30, N 24.55; Found: C 43.63, H 5.17, N 25.73.

 (Z) and (E) $(2R, 3R, 5R)$ -2,3-diazide-5-hydroxycyclohexanone oximes $(27, 28)$. To a solution of 26 (0.14 g, 0.61 mmol) in 96% ETOH (30 mL), NaN₃ (0.5 g, 7.7 mmol) was added. The mixture was refluxed at \sim 40°C. After 2 h TLC (solvent G) showed the end of the reaction. Then the mixture was diluted with $Et₂O (30 mL)$. Precipitated salts were filtered off. The filtrate was concentrated and diluted with AcOEt. The organic solution was washed with water, dried (Na_2SO_4) , and concentrated. The crude product was chromatographed (solvent H) to give first 27 (34%, syrup); $[\alpha]_D^{20} - 47^\circ$ (c 0.1, CHCl₃); R_f 0.50 (solvent H); IR: ν 3200 (OH), 2100 (N₃), 1650 (C=N oxime) cm⁻¹.

Anal. Calcd for $C_6H_9N_7O_2$: C 34.12, H 4.30, N 46.43; Found: C 35.21, H 4.43, N 46.32.

Eluted second was 28 (47%, syrup); $[\alpha]_D^{20} - 25^\circ$ (c 0.4, CHCl₃); R_f 0.48 (solvent H); IR: ν 3240 (OH), 2200 (N₃), 1660 (C=N oxime) cm⁻¹.

Anal. Calcd for $C_6H_9N_7O_2$: C 34.12, H 4.30, N 46.43; Found: C 36.61, H 4.62, N 46.94.

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REFERENCES

- 1. Ferrier, R.J. Unsaturated carbohydrates. Part 21. A carbocyclic ring closure of a hex-5-enopyranoside derivative. J. Chem. Soc., Perkin Trans. 1 1979, 1455–1458.
- 2. Ferrier, R.J.; Middleton, S. The conversion of carbohydrate derivatives into functionalized cyclohexanes and cyclopentanes. Chem. Rev. 1993, 93, 2779–2831.
- 3. Semeria, D.; Philippe, M.; Delaumeny, J.M.; Sepulchre, A.M.; Gero, S.D. A general synthesis of cyclitols and aminocyclitols from carbohydrates. Synthesis 1983, 710–713.
- 4. Chretien, F.; Chapleur, Y. Chiral cyclohexanes from carbohydrates: successful carbocyclization of a D-arabino-hex-5-enopyranoside derivative. J. Chem. Soc., Chem. Commun. 1984, 1268–1269.
- 5. Jung, M.E.; Choe, S.W.T. Total synthesis of cyclophellitol and (1R,2S)-cyclophellitol from D-mannose. J. Org. Chem. 1995, 60, 3280–3281.
- 6. Cleophax, J.; Dubreuil, D.; Gero, S.D.; Loupy, A.; Viera de Almeida, M.V.; Da Silva, A.D.; Vass, G. Enantioselective synthesis of inositols as intermediates for the preparation of deoxy-inositol phosphates from D-galactose. Bioorg. Med. Chem. Lett. 1995, 5, 831–834.
- 7. Dubreuil, D.; Cleophax, J.; Viera de Almeida, M.; Verre–Liaigre, J.; Sebrie´, C.; Vass, G.; Gero, S.D. Stereoselective synthesis of 6-deoxy and 3,6-dideoxy-D-myoinositol precursors of deoxy-myo-inositol phosphate analogues from D-galactose. Tetrahedron 1997, 53, 16747–16766.
- 8. Chung, S.K.; Yu, S.H. Synthesis of L-chiro-inositol-1,2,3-trisphosphate and -1,2,3,5 tetrakisphosphate by Ferrier reaction of methyl α -D-mannopyranoside. Bioorg. Med. Chem. Lett. 1996, 6, 1461–1464.
- 9. Takahashi, H.; Kittaka, H.; Ikegami, S. Novel synthesis of enantiomerically pure natural inositols and their diastereoisomers. Tetrahedron Lett. 1998, 39, 9707–9710.
- 10. Laszlo, P.; Pelyvas, I.F.; Sztaricskai, F.; Szilagyi, L.; Somogyi, A. Novel aspects of the Ferrier carbocyclic ring-transformation reaction. Carbohydr. Res. 1988, 175, 227–239.
- 11. Pelyvas, I.F.; Madi-Puskas, M.; Toth, Z.G.; Varga, Z.; Batta, G.; Sztaricskai, F. Novel aminocyclitol antibiotics derived from natural carbohydrates. Carbohydr. Res. 1995, 272, C5–C9.

- 12. Pelyvas, I.F.; Madi-Puskas, M.; Toth, Z.G.; Varga, Z.; Hornyak, M.; Batta, G.; Sztaricskai, F. Synthesis of new pseudodisaccharide aminoglycoside antibiotics from carbohydrates. J. Antibiot. 1995, 48, 683–695.
- 13. Wang, L.X.; Sakairi, N.; Kuzuhara, H. Synthesis of 1D-(1,3,5/2,4)-4-acetamido-5 amino-1,2,3-cyclohexanetriol and incorporation into a pseudo-disaccharide. Carbohydr. Res. 1995, 275, 33–47.
- 14. Thum, O.; Chen, J.; Prestwich, G.D. Synthesis of a photoaffinity analogue of phosphatidylinositol 3,4-bisphosphate, an effector in the phosphoinositide 3-kinase signaling pathway. Tetrahedron Lett. 1996, 37, 9017–9020.
- 15. Friestad, G.K.; Branchaud, B.P. A new approach to the Pancratistatin C-ring from D-glucose: Ferrier rearrangement, pseudoinversion and Pd-catalyzed cyclization. Tetrahedron Lett. 1997, 38, 5933–5936.
- 16. Amano, S.; Ogawa, N.; Ohtsuka, M.; Chida, N. Chiral and stereoselective total synthesis of novel immunosuppressant FR65814 from D-glucose. Tetrahedron 1999, 55, 2205–2224.
- 17. Amano, S.; Takemura, N.; Ohtsuka, M.; Ogawa, S.; Chida, N. Total synthesis of Paniculide A from D-glucose. Tetrahedron 1999, 55, 3855–3870.
- 18. Imuta, S.; Ochiai, S.; Kuribayashi, M.; Chida, N. New synthesis of (-)- and (+)actinobolin from D-glucose. Tetrahedron Lett. 2003, 44, 5047–5051.
- 19. Hladezuk, I.; Olesker, A.; Tchertanov, L.; Cleophax, J. Preparation of novel polysubstituted chiral cyclohehanone derivatives containing a quaternary carbon by Ferrier reaction. Carbohydr. Res. 2001, 330, 257–265.
- 20. Jaramillo, C.; Chiara, J.L.; Martin-Lomas, M. An effective strategy for the synthesis of $6-O-(2-amino-2-deoxy- α -D-glucopyranosyl)-D-chiro and -D-myo-inositol 1-phos$ phate related to putative insulin mimetics. J. Org. Chem. 1994, 59, 3135–3141.
- 21. Dabrowska, A.; Smiatacz, Z. Synthesis and structure of some methyl 2,3-dideoxy-3azido-D-hexopyranosides. In Book of Abstracts, VIIth European Carbohydrate Symposium, Cracow, Poland, August 22–27, 1993; A015.
- 22. Dabrowska, A.; Konitz, A.; Smiatacz, Z. Synthesis, the crystal structure, and highresolution NMR spectroscopy of methyl 4-O-acetyl-3-azido-2,3,6-trideoxy-6-iodo- α -D-arabino-hexopyranoside. Carbohydr. Res. 2002, 337, 175–181.
- 23. Liberek, B.; Da˛browska, A.; Frankowski, R.; Matuszewska, M.; Smiatacz, Z. The use of tri-O-acetyl-D-glucal and -D-galactal in the synthesis of 3-acetamido-2,3-dideoxyhexopyranoses and -hexopyranosides. Carbohydr. Res. 2002, 337, 1803–1810.
- 24. Lemieux, R.U.; Georges, F.Z.G.; Smiatacz, Z. A novel approach to the synthesis of a variety of aminodeoxy- α -D-glycopyranosides. Can. J. Chem. 1981, 59, 1433–1438.
- 25. Smiatacz, Z.; Szweda, R.; Myszka, H. The structure and properties of the products of reaction between 3,4,6-tri-O-acetyl-2-deoxy-2-nitroso-a-D-galactopyranosyl chloride and pyrazole. Carbohydr. Res. 1986, 153, 33–43.
- 26. Liberek, B.; Frankowski, R.; Smiatacz, Z. Synthesis and properties of derivatives of 2-deoxy-2-hydroxyimino-D-arabino-hexopyranosyl-L-cysteine and thiophenol. Polish J. Chem. 1999, 73, 1153–1167.
- 27. Garegg, P.J.; Samuelsson, B. Novel reagent system for converting a hydroxy-group into iodo-group in carbohydrates with inversion of configuration. J. Chem. Soc., Perkin Trans. 1 1980, 2866–2868.
- 28. Whistler, R.L.; Kashem, A.; Anisuzzaman, A.K.M. Preferential halogenation of the primary hydroxyl group. Meth. Carbohydr. Chem. 1980, 31, 227–231.
- 29. Liberek, B.; Sikorski, A.; Melcer, A.; Konitz, A. X-ray diffraction and high-resolution NMR spectroscopy of methyl 3-azido-2,3-dideoxy- α -D-lyxo-hexopyranoside. Carbohydr. Res. 2003, 338, 795–799.
- 30. Machado, A.S.; Dubreuil, D.; Cleophax, J.; Gero, S.D.; Thomas, N.F. Expedient syntheses of inososes from carbohydrates: conformational and stereoelectronic aspects of the Ferrier reaction. Carbohydr. Res. 1992, 233, C5–C8.
- 31. Lemieux, R.U.; Earl, R.A.; James, K.; Nagabhushan, T.L. Isopropyl tri-O-acetyl- α -Dhexopyranosulosides. The configurations of the parent oximes. Can. J. Chem. 1973, 51, 19–26.
- 32. Liberek, B.; Konitz, A.; Frankowski, R.; Smiatacz, Z. The synthesis and structure of the derivatives of 2-deoxy-2-hydroxyimino-D-lyxo-hexopyranosyl-L-cysteine and -thiophenol. Carbohydr. Res. 2000, 326, 151–158.

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